Collaboration between clinicians, geneticists, and basic scientists as well as the clever use of mouse models will certainly help address some of these questions and enhance our understanding of IRF6- and p63-related diseases, the ultimate beneficiaries being the afflicted patients and their families.

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C. neoformans is a soil-dwelling fungus that emerged in the late twentieth century as a major human pathogen because of its propensity to cause lethal meningocerebralitis. The burden of cryptococcosis is estimated to approach one million cases per year, with a mortality that exceeds that from tuberculosis (1). C. neoformans is acquired by inhalation of dehydrated cells or spores (2). Serologic surveys indicate a high prevalence of human infection, which is likely to be first acquired in childhood (3). Although infection is common, disease is rare, and cryptococcosis occurs primarily in hosts with impaired immunity, such as patients with AIDS, organ transplant recipients, and those treated with immunosuppressive therapies (2). Hence, normal immune responses are believed to control infection in the lung. Extrapulmonary dissemination is therefore invariably associated with disease, with meningocerebralitis being the most common clinical presentation of cryptococcosis. To cause meningocerebralitis, C. neoformans must cross several epithelial and/or endothelial cell layers, first to leave the lung and then to reach the brain. How does a soil-dwelling organism that has no need for animal pathology reach the brain to cause meningoencephalitis? In this issue of the JCI, Shi et al. use intravital microscopy to reveal that brain invasion by C. neoformans follows a capillaric microembolic event. They find that after suddenly stopping in brain capillaries, cryptococci cross into the central nervous system in a process that is urease dependent, requires viability, and involves cellular deformation. This observation provides evidence for direct brain invasion by C. neoformans, but a consideration of all the currently available evidence suggests a role for both direct and phagocyte-associated invasion. Hence, the remarkable neurotropism of C. neoformans may have more than one mechanism.

Cryptococci at the brain gate: break and enter or use a Trojan horse?
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The mechanism by which Cryptococcus neoformans invades the central nervous system is fundamental for understanding pathogenesis because cryptococcosis commonly presents as meningocerebralitis. There is evidence for both direct invasion of the endothelial cells lining the brain vasculature and a “Trojan horse” mechanism whereby cryptococci enter the central nervous system after macrophage ingestion. However, in this issue of the JCI, Shi et al. use intravital microscopy to reveal that brain invasion by C. neoformans follows a capillaric microembolic event. They find that after suddenly stopping in brain capillaries, cryptococci cross into the central nervous system in a process that is urease dependent, requires viability, and involves cellular deformation. This observation provides evidence for direct brain invasion by C. neoformans, but a consideration of all the currently available evidence suggests a role for both direct and phagocyte-associated invasion. Hence, the remarkable neurotropism of C. neoformans may have more than one mechanism.
the technique of intravital microscopy to visualize in mice the process of brain invasion by *C. neoformans*.

**Current views of brain invasion**

Although the predisposition of *C. neoformans* to cause meningoencephalitis has been known for more than a century, the mechanism by which fungal cells invade the central nervous system has remained elusive. In recent years, two competing hypotheses have been proposed for brain invasion (Figure 1). The first mechanism posits a “Trojan horse” approach, whereby fungal cells gain access to the brain by transport in phagocytic cells. The finding that cryptococci in the meningeal vasculature were in close association with phagocytic cells suggested that brain invasion was cell associated (5). Circumstantial evidence for this mechanism is provided by the fact that *C. neoformans* is a facultative intracellular pathogen that can survive in macrophages (6) and that extrapulmonary dissemination appears to be macrophage associated (7–9). Strong experimental evidence for the Trojan horse mechanism came from elegant experiments in which mice were infected with macrophages containing ingested cryptococci (10). According to this view, fungal cells are phagocytosed first in the blood or the vicinity of the endothelial cells of the brain vasculature and then the phagocytic cell transports them to the parenchyma. The second mechanism posits that naked *C. neoformans* cells invade the brain by direct transcytosis of endothelial cells lining the brain vasculature (11). This view is supported by in vitro and in vivo observations showing that yeast cells are taken up by endothelial cells and can transit through the cytoplasm to emerge on the other cellular surface (11). It is noteworthy that neither mechanism is exclusive of the other, and in fact, there is some evidence that both can occur simultaneously (10).

**Intravital microscopy provides new insights**

In this issue of the *JCI*, Shi et al. (4) apply the technique of intravital microscopy to visualize in mice the process of brain invasion by *C. neoformans*. They were able to visualize the brain microvasculature adjacent to the meninges and observe the events preceding, and concurrent with, cryptococcal brain invasion. A major finding of this study is that fungal cells stop suddenly in capillary sites without rolling or tethering, in a manner similar to polystyrene microspheres. The similarity of the arresting process to that exhibited by inert microspheres suggests that the initial brain localization mechanism is mechanical and related to an inability of yeast cells to traverse narrow capillaries. If this is the case, the initial brain infection results from a microembolic event. It was observed that after stopping, *C. neoformans* cross the capillary wall in a process that requires viability but not replication, is associated with deformation of cell morphology, and is urease dependent, as reported previously (12). Finally, the investigators show that inhibiting urease reduces brain fungal burden, suggesting that this might provide an entirely new approach toward protecting the brain in cryptococcal meningitis. Each of these observations has important repercussions for our understanding of cryptococcal neuropathogenesis.

The finding that the initial brain localization followed sudden arrest in what appears to be a fungal microembolic event suggests that the process may not require specific attachment receptors, as
has been suggested by in vitro studies (13, 14), although these receptors could still play a role in invasion. If this is the case, the remarkable neurotropism of this fungus may have more to do with mechanical entrainment than with either microbi-specific neurotrophic properties or the immune-privileged status of the central nervous system. However, mice inoculated with encapsulated and non-encapsulated strains of C. neoformans manifest comparable brain infection (4), a finding that argues against an effect based solely on diameter, since one might have expected more brain invasion after injection of the encapsulated cells, which are larger by virtue of having a capsule. A caveat in this comparison is that acapsular cells have a propensity to aggregate, and thus it is conceivable that the size difference between encapsulated and acapsular cells was not apparent because the latter involved aggregates of more than one cell. It would be interesting to know whether other yeast cell types such as Candida albicans or Saccharomyces cerevisiae also manifest sudden stopping in the brain vasculature. If these species behave as C. neoformans, then the remarkable neurotropism of cryptococci must have another explanation, since candidemia is not associated with meningoencephalitis. Furthermore, the contribution of cell size and capsule to initial brain localization could also be explored by comparing cells with induced and non-induced capsules. If mechanical trapping is all that is needed for initial invasion, then one might predict that intravenous infection with the larger capsule-induced cells should produce a greater burden of brain infection than infection with smaller cells. This, in turn, would imply a new pathogenic function for the capsule.

Shi et al. (4) also observed C. neoformans directly crossing the capillaries into the brain parenchyma following sudden stopping. Although this observation provides direct in vivo experimental support for the transcytosis model of cryptococcal brain invasion (Figure 1), it is worthwhile pausing and considering the advantages and limitations of this system in the interpretation of the data. The major advantage of intravenous infection is that it allows the researcher to control the time of blood invasion so that real-time visualization of the pathogenic process is possible. However, intravenous infection may or may not reflect the natural mode of brain dissemination. There is widespread consensus in the field that the initial site of human infection is the lung and that extrapulmonary dissemination occurs only in hosts whose immune systems cannot control the fungus in the lung. Macrophages appear to be critically important cells in determining the outcome of infection, such that resistance to infection has been correlated with their ability to restrict intracellular fungal replication (15). Recently, indirect evidence has emerged that extrapulmonary dissemination is cell associated, with fungal cells leaving the lung in phagocytic cells, possibly macrophages. If this is the case, then the intravenous model, whereby fungal cells are injected directly into the bloodstream, may not seem that relevant to the natural processes responsible for meningoencephalitis. However, even if extrapulmonary dissemination is cell associated, the finding that C. neoformans is capable of nonlytic exocytosis of infected cells (16, 17) suggests that infected cells can release free C. neoformans into the circulation that would then be in a situation analogous to that following intravenous infection (16, 17). Another caveat in interpreting the results of Shi et al. (4) is that cryptococci are normally ingested by macrophages in chronic pulmonary infection, and the current experiment was done in mice infected intravenously such that there may not have been sufficient time for phagocytic cells to ingest them. Repeating the experiments performed by Shi et al. (4) using a pulmonary route of infection with C. neoformans might provide data to resolve the direct versus Trojan horse invasion scenarios, although such studies could be technically demanding, since the researcher would not be able to control the timing of extrapulmonary dissemination and know when to monitor the brain vasculature. However, perhaps this experimental hurdle could be bypassed by injecting macrophages containing ingested cryptococci and observing their interaction with the brain vasculature.

Fungal activities promote invasion

In the study by Shi et al. (4), there are other pearls worthy of comment. First, the observation that the fungus changes its shape from spherical to ovoid during the process of brain invasion, which raises the possibility that the cryptococcal cell skeleton and cell wall are altered to facilitate this process. This observation is fascinating, because it implies that C. neoformans, like other fungi, is capable of morphological change, in a process that gives it access to another niche and has distant echoes in the morphological changes observed during tissue invasion by dimorphic fungi and C. albicans. Hence, shape changes accompany the changes in capsule structure that have been described during brain invasion (18). Second, the finding that urease activity contributes to brain infection is an observation that merits intensive study to ascertain the mechanisms involved. Third, the observation that pharmacologic inhibition of urease activity reduces brain infection provides a new precedent for protecting the brain. Although such an approach is likely to have limited usefulness in most cases of human cryptococcosis, where the majority of patients present with meningoencephalitis, and therefore already have substantial brain infection, the notion that enzymatic inhibition translates into reduced neural invasion does open new areas of investigation that may be exploited in the future. For example, if the effect of urease is extracellular, it might be possible to protect the brain parenchyma against cryptococcal infection by raising urease-specific neutralizing antibody responses.

Transmigration and/or Trojan horses

The article by Shi et al. (4) is a major contribution to the cryptococcal field because it applies a new technique to the study of the pathogenesis of C. neoformans meningoencephalitis that gives fresh new insights into the mechanisms involved and pose many new questions for future studies. As to the question of whether brain invasion is the result of direct transmigration or occurs via a Trojan horse, while the data generated by Shi et al. (4) support the former, the totality of the available evidence strongly suggests that both mechanisms occur. Debates as to the relative importance of each mechanism are not helpful, since it is likely that one or the other predominates depending on the host and/or the conditions of the experiment. Instead, it is fortunate that the cryptococcal field has at least two different mechanisms for brain invasion to study, each fascinating on its own, and each providing an abundance of questions for future study.

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The mammalian target of rapamycin (mTOR) signaling pathway is activated in several disorders associated with benign tumors and malformations of the cerebral cortex. In this issue of the *JCI*, Orlova et al. have now definitively added another disorder to this group by demonstrating that activation of mTOR signaling is associated with polyhydramnios, megalencephaly, and symptomatic epilepsy syndrome (PMSE), which is characterized by severe intractable epilepsy and megalencephaly. PMSE is caused by lack of the pseudokinase STE20-related kinase adaptor α (STRADα), and Orlova et al. show that reduction of STRADα levels during corticogenesis in the mouse results in a cellular phenotype and neuronal migration defects similar to those observed in patients with PMSE, clearly demonstrating a pivotal role for STRADα in cell polarity and growth. This study helps pave the way for possible therapeutic intervention with rapamycin to control the epilepsy and learning disabilities associated with this disorder.

Development of the human cortex is a complex and finely orchestrated process. The layers of the developing neocortex are generated through neuronal migration, whereby neurons travel from their place of origin in the proliferating ventricular zone to their final position in the brain. This cortical lamination is generated from the inside out, with the neurons that contribute to each layer traveling past the existing layers; perturbation of this carefully controlled neuronal migration can result in heterotopia (neurons in the wrong place) and dysplasia (disorganization of the normal structure of the cortex) that disrupt the normal cortical circuitry. Imbalance between excitatory and inhibitory systems in the cortex can lead to spontaneous electrical discharge with catastrophic consequences, often in the form of intractable or medication-resistant epilepsy and severe intellectual disability (1).

Polyhydramnios, megalencephaly, and symptomatic epilepsy syndrome (PMSE), recently described in an Old Order Mennonite population, is associated with craniofacial dysmorphism (large forehead, widely spaced eyes, and large mouth), an abnormally large brain, and severe, early-onset intractable epilepsy (2). PMSE in this kindred is caused by homozygous deletion of a portion of the STE20-related kinase adaptor α gene (STRADA; encoding STRADα) on human chromosome 17 (2). STRADα normally binds and exports the protein kinase serine/threonine kinase 11 (STK11; also known as LKB1) out of the nucleus, where they bind to MO25 to form a trimeric complex that has an inhibitory effect on mammalian target of rapamycin (mTOR) signaling through the sequential phosphorylation of AMPK and the tuberous sclerosis complex 2 (TSC1/TSC2) complex (Figure 1 and ref. 3). Aberrant activation of the mTOR pathway was identified in the brain of an individual with PMSE by the presence of high levels of phosphorylated ribosomal S6 protein (2), a downstream target of mTOR. This finding is suggestive of a link between the cellular mechanism underlying this disorder and that previously shown to be responsible for a group of syndromes characterized by hamartomas, benign tumors composed of differ-