Translation of ASH1 mRNA is repressed by Puf6p–Fun12p/eIF5B interaction and released by CK2 phosphorylation

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Translational repression during mRNA transport is essential for spatial restriction of protein production. In the yeast Saccharomyces cerevisiae, silencing of ASH1 mRNA before it is localized to the bud cortex in late anaphase is critical for asymmetric segregation of Ash1p to the daughter cell nucleus. Puf6p, an ASH1 mRNA-binding protein, has been implicated in this process as a translational repressor, but the underlying mechanism is unknown. Here, we used yeast extract-based in vitro translation assays, which recapitulate translation and phosphorylation, to characterize the mechanism of Puf6p-mediated translational regulation. We report that Puf6p interferes with the conversion of the 48S complex to the 80S complex during initiation, and this repression by Puf6p is mediated through the general translation factor eIF5B (Fun12p in S. cerevisiae). Puf6p interacts with Fun12p via the PUF domain, and this interaction is RNA-dependent and essential for translational repression by Puf6p. This repression is relieved by phosphorylation of the N-terminal region of Puf6p mediated by protein kinase CK2 (casein kinase II). Inhibition of phosphorylation at Ser31, Ser34, and Ser35 of Puf6p increases its translational repression and results in ASH1 mRNA delocalization. Our results indicate that Puf6p suppresses the translation initiation of ASH1 mRNA during its transport, and this repression can be released by CK2 phosphorylation in the N-terminal region of Puf6p when the mRNA reaches the bud tip.

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tip (Gonzalez et al. 1999; Irie et al. 2002). A casein kinase I (CK1) protein kinase-mediated release of the translational control by Khd1p has been identified recently [Paquin et al. 2007]. The mechanism by which Puf6 functions as a translational repressor and how this repression is released remain elusive.

In this study, we examine the role of Puf6p, a PUF protein, in regulating translation of ASH1 mRNA. We show that Puf6p represses translation by interfering with the conversion of the 48S complex to 80S, and that this repression is mediated through the general translation initiation factor elf5B/Fun12p. Both the N-terminal region and the PUF domain of Puf6p are required for Puf6p repression activity. Casein kinase II (CK2) phosphorylation sites on Puf6p have been identified in the N-terminal region, and CK2 phosphorylation reduces Puf6p repression activity. CK2 localizes to the bud tip before ASH1 expression. These results suggest a mechanism of translational repression by Puf6p involving Fun12p and a spatially controlled phosphorylation step to relieve it.

Results

Puf6p interferes with 80S assembly in translation initiation

To investigate the mechanism of translation regulation by Puf6p, we developed an in vitro translation assay using cell-free yeast extracts. We constructed a reporter mRNA with the coding sequence for renilla luciferase and a 3′ untranslated region (UTR) containing the E3 element of ASH1 mRNA (15 nucleotides [nt] of the coding sequence of ASH1 mRNA and 121 nt of the 3′UTR) that has been shown to be recognized by Puf6p [Fig. 1A; Gu et al. 2004]. A similar construct lacking the E3 element (R-luc) serves as a control for specificity. We incubated the in vitro synthesized mRNA with yeast extracts containing [35S]methionine and verified the synthesis of renilla protein (data not shown). We preincubated this reporter mRNA with recombinant Puf6 and measured its translational activity. In the presence of Puf6, considerably less protein was produced using R-luc mRNA than with R-luc-E3 mRNA [Fig. 1B], by 82% and 94% at protein-to-RNA molar ratios of 20 and 100, respectively. The renilla luciferase synthesized from R-luc mRNA without the Puf6p-binding site (E3 element) did not show a dosage-dependent decrease in the presence of Puf6. Thus, Puf6 represses translation in a sequence-specific fashion in yeast extracts, consistent with the results obtained using rabbit reticulocyte lysates [Gu et al. 2004].

Translation initiation is the rate-limiting step in translation and serves as a target for translational regulation (Dever 2002). To dissect the mechanism of Puf6p inhibition, we analyzed the distribution of translation complexes by sucrose density gradient. To increase the resolution, we constructed a short coding region followed by the E3 element of ASH1, 179 nt in total [Trachsel et al. 1977]. Cycloheximide was used to inhibit translation elongation. We found that the labeled E3 RNA under control conditions sedimented at fractions 9–11 [Fig. 1C, filled circles], which corresponded to the 80S complex from the absorbance profile (Supplemental Fig. 1), indicating that the reporter was translated. In the presence of GMP-PNP, a nonhydrolyzable analog of GTP that blocks 60S subunit joining [Lee et al. 2002], the E3 RNA formed a complex sedimenting in fractions 3–5 that corresponded to the 48S complex [Fig. 1C, open circles]. The decrease of the 80S complex and concomitant increase of the 48S complex indicated that GMP-PNP allowed translation initiation to proceed to the 48S complex but effi-
Fun12p associates with Puf6p and is required for ASH1 translation and localization

General translation initiation factors are common targets for translational regulation. Puf6p was found to interact with Fun12p [eIF5B in yeast], which assists 60S subunit joining (Gavin et al. 2002, Lee et al. 2002, Collins et al. 2007). To evaluate the possibility that Puf6p could repress translation initiation by suppressing Fun12p, we first tested the interaction between Puf6p and Fun12p by communoprecipitation (co-IP) using yeast extracts from cells expressing both Fun12p-HA and Puf6p-Tap. Fun12p-HA coprecipitated from cells expressing Puf6p-Tap but did not from cells with untagged Puf6p using Tap purification (Fig. 2A). In the reciprocal co-IP, we found Puf6p-HA coprecipitated with Fun12p-Tap but not with untagged Fun12p (Fig. 2B). This interaction is RNA-dependent, as the association between Puf6p and Fun12p was significantly decreased after RNase treatment (Supplemental Fig. 2A).

Although Fun12p is a general translation initiation factor, it is not essential for cell viability (Choi et al. 1998). To test if Fun12p was required for ASH1 translation, we disrupted FUN12 in a strain expressing Ash1p-Myc (fun12). The protein concentrations of Ash1, Pgk1, and Nop1p were reduced by deletion of FUN12. However, the reduction of Ash1p was considerably more significant than Pgk1p and Nop1p whose mRNAs do not contain consensus Puf6p-binding sites (Fig. 2C). We found the expression of Ash1p decreased by >80% relative to Pgk1p or Nop1p after the disruption of FUN12 (Fig. 2D). ASH1 mRNA levels were reduced by 50% relative to ACT1 in the fun12 strain (Fig. 2E), suggesting that both transcription and translation of ASH1 mRNA were reduced by FUN12 disruption. An GST-Fun12 fusion recombinant protein containing amino acid residues 396–1002 was functional in vivo, as it was able to fully complement the slow-growth phenotype of the fun12Δ strain and restore protein synthesis in vitro (Choi et al. 1998, 2000). Truncated Fun12p complemented the expression of Ash1p in a fun12Δ strain when introduced on a shuttle plasmid. Incubation of the E3 RNA with recombinant Puf6p resulted in an increase in the 48S complex and a decrease in the 80S complex similar to results obtained with GMP-PNP (Fig. 1C, filled triangles). This increase of the 48S peak was reproducible using different gradients [see the Materials and Methods]. This suggested that assembly of the 80S complex but not the formation of the 48S complex was affected by Puf6p. Inhibition by Puf6p was specific, as competition with cold E3 RNA led to a recovery of the 80S complex preformed by the labeled E3 (Fig. 1C, open and filled squares), consistent with the essential role of Mg2+ in RNA-ribosomal complex formation. These results suggest that Puf6p blocked 80S complex assembly but not 48S complex formation during translation initiation, which may result from its effect on 60S joining.

**Figure 2.** Fun12p associates with Puf6p and is required for ASH1 mRNA translation and localization. (A) Co-IP with protein A tag [Tap] and IgG-coated beads. Proteins in yeast extract expressing Fun12p-HA [lanes 1,3] or Puf6p-Tap/Fun12p-HA [lanes 2,4] were affinity-purified on matrix-bound IgG. Eluates [lanes 3,4] and 16% total inputs [lanes 1,2] were analyzed by Western blotting with anti-HA antibody. (B) Reciprocal co-IP of Puf6p and Fun12p. Inhibition by Puf6p was specific, as competition with cold E3 RNA led to a recovery of the 80S complex preformed by the labeled E3 RNA (Fig. 1C, open and filled triangles). In the presence of EDTA, RNA–ribosomal complexes were not detected, independent of Puf6 (Fig. 1C, open and filled triangles). This increase of the 48S peak was reproducible using different gradients [see the Materials and Methods]. This suggested that assembly of the 80S complex but not the formation of the 48S complex was affected by Puf6p. Inhibition by Puf6p was specific, as competition with cold E3 RNA led to a recovery of the 80S complex preformed by the labeled E3 RNA (Fig. 1C, open and filled triangles). This increase of the 48S peak was reproducible using different gradients [see the Materials and Methods].
a plasmid (Supplemental Fig. 2B). Therefore, this truncated Fun12p could form the initiation complex on ASH1 mRNAs, as does the full-length Fun12p in vivo. FUN12 overexpression increased the concentration of Ash1p by twofold in comparison with Pgk1p but had only a marginal difference at the mRNA level (Supplemental Fig. 3), consistent with the requirement for Fun12p in ASH1 mRNA translation.

Although fun12 cells grow slower than wild type (Choi et al. 1998), the cell morphology of fun12 has been shown to be normal (Narayanaswamy et al. 2006) and phalloidin staining displayed normal actin organization (data not shown). We then analyzed the effect of FUN12 disruption on ASH1 mRNA localization. ASH1 mRNA was delocalized in the fun12 mutant (Fig. 2F). In the wild-type strain, 80% of ASH1 mRNA was localized at the distal cortex of the bud. However, in the fun12 mutant, ASH1 mRNA was localized diffusely within the bud (62%), in the neck (16%), or in the mother and bud (12%) (Fig. 2G). This pattern of ASH1 mRNA in fun12 cells was different from the she mutants where the ASH1 mRNA was mostly diffused throughout mother and bud (Long et al. 1997). To test whether Fun12p was also required for the localization of other mRNAs, we examined the localization of IST2 mRNA (Takizawa et al. 2000), which contains putative Puf6p-binding sites at the 3'UTR (Gu et al. 2004). IST2 mRNA was found diffusely distributed within the bud in both fun12 and puf6 mutants (Supplemental Fig. 4), similar to the pattern observed for ASH1 mRNA.

Both the N-terminal region and PUF domain of Puf6p are required for translational repression

Puf6p contains a PUF domain and a distinct N-terminal region with low homology compared with other PUF proteins (Supplemental Fig. 5). We generated two fragments of Puf6 to dissect their functional roles [Fig. 3A]: N120 (N-terminal region outside of the PUF domain), and C536 (PUF domain plus C terminus). Since the PUF domain (amino acids 121–565) was insoluble when expressed in Escherichia coli, we designed a fragment (C536) containing both the PUF domain and the C terminus of Puf6p (amino acids 121–656). The recombinant proteins of full-length, N120, and C536 with N-terminal His6 tag were purified from E. coli (Fig. 3B). N120 showed a higher molecular weight (~33 kDa) than expected (18.7 kDa) by SDS-PAGE. This might be due to insufficient binding of SDS to the Asp/Glu-rich region in the N-terminal region of Puf6p (Gu et al. 2004).

We tested RNA-binding activity of these three Puf6 recombinant proteins to E3 RNA using a polyacrylamide gel electrophoretic assay. Both C536 and full-length Puf6 bound to the E3 RNA and resulted in a band shift [Fig. 3C], consistent with the finding that PUF domain is an RNA-binding motif (Zamore et al. 1997). N120 showed a weaker shift with the RNA, although it does not contain a known RNA-binding motif.

We next examined the interaction of the three forms of recombinant Puf6 with Fun12p by pull-down assays. The recombinant Puf6 were incubated with yeast extract containing Tap-tagged Fun12p and purified on nickel beads. We found Fun12p-Tap was retained by C536 and full-length Puf6, but not by N120 (Fig. 3D), suggesting that the PUF domain is required for the interaction with Fun12p. This result also confirms the interaction of Puf6p and Fun12p from the co-IP studies.

Since C536 interacts with both Fun12p and RNA, it could be sufficient to repress translation of E3 RNA. To test this, we performed in vitro luciferase translation assays using yeast extract and different forms of Puf6. As shown in Figure 3E, in comparison with R-luc RNA, full-length His6-Puf6 repressed translation of R-luc-E3 RNA specifically by half. This repression was comparable with the results using the PhosphorImager (Fig. 1B), confirming luciferase assays as a quantitative measure of in vitro translated protein. In contrast, C536 repressed both R-luc-E3 and R-luc RNA equivalently by 20% in comparison with the reaction without recombinant proteins [Fig. 3E], indicating that the PUF domain alone was insufficient for E3-specific translational repression. Repression by N120 was not observed for either R-luc-E3 or
Puf6p is phosphorylated by protein kinase CK2

One role for N120 could be as a regulatory domain for Puf6p-mediated repression. Cka2p, a catalytic subunit of protein kinase CK2, was also detected in the She2p-Tap affinity purification together with Puf6p (Gu et al. 2004, data not shown). Global protein interaction studies showed that Puf6p associated with each of the two catalytic subunits of CK2 (Ho et al. 2002), suggesting that Puf6p may be a substrate for CK2. Relevant to this, we found six potential CK2 phosphorylation sites in N120 region but only one in the C536 region.

We first tested whether Puf6p was phosphorylated in vivo using a sensitive noncovalent fluorescent dye staining technology for the detection of phosphoserine-, phosphothreonine-, and phosphotyrosine-containing proteins displayed on SDS-PAGE (Pro-Q diamond phosphoprotein gel staining, Molecular Probes). The Puf6p-Tap purified from yeast extracts was found to be phosphorylated (Fig. 4A). The phosphorylation of Puf6p was specific as the untagged cell did not show a phosphoprotein band with the similar gel mobility (Supplemental Fig. 6). The phosphorylation of endogenous Puf6p was further confirmed by treatment with λ phosphatase (λ pPase), which can remove phosphates from both Ser/Thr and Tyr. After λ pPase treatment, a faster-migrating band for Puf6p-Tap was detected (Fig. 4B), suggesting phosphate groups were removed from Puf6p. The λ pPase treatment also caused a faster migration of Puf6p-HA purified from yeast, confirming that the phosphorylation is on Puf6p and not the tag (Fig. 4B). Only a single band of Puf6p was detected by Western blot before the treatment of λ pPase, suggesting that the majority of endogenous Puf6p may be phosphorylated.

To test which region of Puf6p was phosphorylated, we performed an in vitro phosphorylation assay with recombinant Puf6p proteins. Full-length Puf6 and N120, but not C536, were phosphorylated when incubated with yeast extracts (Fig. 4C). A similar pattern was observed in the phosphorylation assay using CK2 purified from sea star (Fig. 4D), suggesting that phosphorylation of His6-Puf6 in yeast extract is likely due to endogenous CK2. A special feature of CK2 is that it can use both ATP and GTP as phosphate donors (Gatica et al. 1993). We found that yeast extract phosphorylated full-length Puf6 and N120 but not C536 in the presence of [γ-32P]-GTP or [γ-32P]-ATP (Fig. 4E). The phosphorylation of Puf6p by yeast CK2 was further confirmed by the finding that recombinant Puf6 was efficiently phosphorylated by yeast CK2.

Figure 4. Puf6p is phosphorylated by CK2 on the N-terminal region. (A) Yeast Puf6p is phosphorylated. Proteins in yeast extract from cells expressing Puf6p-Tap or Fun12p-Tap were affinity-purified on matrix-bound IgG. The IgG bead eluates were analyzed by Pro-Q Diamond gel staining for phosphoprotein (column 1), then SYPRO Ruby gel staining for total protein (column 2), and then processed for immunostaining with anti-Tap antibody (column 3). Fun12p-Tap was used as a negative control for phosphoprotein stain. (P-Marker) Phosphoprotein marker. (B) Proteins in yeast extract from cells expressing Puf6p-Tap or Puf6p-HA were affinity-purified and subjected to λ pPase assay. Eluates were analyzed by Western blotting with anti-HA and anti-Tap antibodies. Puf6p-Tap and Puf6p-HA were precipitated by IgG and anti-HA antibody-coupled beads, respectively. IgG light chain was used as an internal control for the gel mobility shift. (C) In vitro phosphorylation assay with yeast extract and [γ-32P]-ATP. Proteins were sampled as indicated during incubation and analyzed by autoradiograph (left) and Western blotting (right). (D) Full-length recombinant proteins; (N) N120 recombinant proteins; (C) C536 recombinant proteins. (D) In vitro phosphorylation assay with CK2 from sea star and [γ-32P]-ATP. Proteins were analyzed by autoradiograph (left) and Coomassie blue staining (right), respectively. (E) In vitro phosphorylation assay with yeast extract and [γ-32P]-ATP or [γ-32P]-GTP as above. Proteins were analyzed by autoradiograph. (F) Mass spectra of the tryptic peptides from unphosphorylated [panel a] and phosphorylated His6-Puf6 [panel b]. In panel a, the species observed at 833.9 m/z corresponds to the doubly charged ion of the peptide ISIDSSDEESELSK (1665.74 Da). The ion that has a mass of 80 higher than ISIDSSDEESELSK is doubly charged, which is expected for addition of two phosphate groups (80 Da for one phosphate group) on the ion.
purified from cells expressing Cka2p-Tap [Supplemental Fig. 7].

To identify the phosphorylation sites, the recombinant full-length His₆-Puf6 was phosphorylated in yeast extract and subjected to mass spectrometry analysis. Two phosphorylation sites, Ser34 and Ser35, were identified [Fig. 4F]. This was consistent with the result from a recent proteomic study that two phosphorylated serines (Ser35 and either Ser31 or Ser34) were identified on endogenous Puf6p (Chi et al. 2007). Ser34 and Ser35 (within 3'DSSSDE39) are canonical CK2 sites: S/T-XX-D/E (X for any nonbasic amino acid) (Meggio and Pinna 2003), and Ser31 could be primed to become a CK2 site by phosphorylation on Ser34 [Roach 1991]. The agreement of the mass spectrometry with the results from in vitro phosphorylation assays strongly suggests that Puf6p is phosphorylated by CK2 on the N-terminal region in vivo, and yeast extract competent for translation also recapitulates this phosphorylation on recombinant Puf6.

**Figure 5.** CK2 phosphorylation relieves translational repression

To test whether phosphorylation of Puf6p affects its function in regulating translation, we generated Ser-to-

 purined from cells expressing Cka2p-Tap [Supplemental Fig. 7].

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**Figure 5.** CK2 phosphorylation relieves translational repression by Puf6p. (A) In vitro phosphorylation assay with rat CK2 and [γ-32P]-ATP. Wild-type [Wt] and Ser31,34,35Ala mutant [Mut] His₆-Puf6 were analyzed by autoradiograph and Coomassie blue staining. (B) Luciferase assay for R-luc-E3 in vitro translated in yeast extract. RNAs were preincubated with wild-type [Wt] or Ser31,34,35 → Ala mutant [Mut] His₆-Puf6. Luciferase activities were normalized to reaction with wild-type His₆-Puf6 at a protein-to-RNA molar ratio of 18. Protein-to-RNA molar ratio varied with [RNA] = 17 nM (X-axis). (C) In vitro phosphorylation assay of Puf6 with rat CK2 or yeast extract and [γ-32P]-ATP with or without DMAT (25 µM). Wild-type [Wt] and Ser31,34,35Ala mutant [Mut] His₆-Puf6 were analyzed by autoradiograph. (D) Luciferase assay for R-luc-E3 and R-luc in vitro translated in yeast extract. RNA was preincubated with wild-type His₆-Puf6 at protein-to-RNA molar ratio of 36 with [RNA] = 17 nM. Luciferase activity was normalized to reaction without DMAT for each reporter. [E–G] Phosphorylation of Puf6 reduces RNA binding. (E) RNA-binding assay with 32P-labeled miniORF-E3 RNA and Puf6p-Tap affinity-purified from yeast extract. Extracts containing Puf6p-Tap or untagged Puf6p were incubated with IgG-coated beads and immobilized proteins were treated or not treated with λ ppase before the RNA-binding assay. RNA-binding activity was normalized to the Puf6p-Tap extract without λ ppase treatment. (F) RNA-binding assay with 32P-labeled miniORF-E3 RNA and Puf6p-Tap affinity-purified from yeast extract as described above. The two UUGU sequences essential for Puf6p binding were deleted in E3 [miniORF-E3-M]. RNA-binding activity was normalized to the Puf6p-Tap extract without λ ppase treatment. (G) RNA-binding assay with 32P-labeled miniORF-E3 RNA and recombinant His₆-Puf6 immobilized on nickel beads. Recombinant His₆-Puf6 was phosphorylated with rat liver CK2 [–ATP] or not [–ATP] before the RNA-binding assay. RNA-binding activities were normalized to binding assays without recombinant Puf6.
the affinity of Puf6p for RNA. To test this possibility, we performed an RNA-binding assay using endogenous Puf6p treated with λ ppase. Puf6p-tap was affinity-purified on matrix-bound IgG from yeast extracts and treated with λ ppase. We then incubated the protein with 32P-labeled E3 RNA. The RNA bound to the IgG beads was eluted and quantified by radioactivity. We found that the labeled E3 RNA bound to the IgG beads was specifically retained by Puf6p-Tap and the retention increased after the treatment with λ ppase [Fig. 5E]. The binding specificity was tested by a mutant E3 RNA that contained mutations in the elements essential for Puf6p binding [Gu et al. 2004]. In comparison with wild-type E3, the mutant E3 RNA was retained less efficiently by Puf6p-Tap [Fig. 5F]. In addition, binding of the mutant E3 was not changed after λ ppase treatment [Fig. 5F], suggesting that the effect of phosphorylation on RNA binding was specific for Puf6p. We also found that E3 RNA bound less efficiently to Puf6 phosphorylated by purified CK2 than unphosphorylated Puf6 [Fig. 5G]. These results suggest that phosphorylation of Puf6p by CK2 can reduce RNA binding and thus affect the repressing activity of Puf6p, similar to the role of phosphorylation in regulating Khd1p, another translational repressor [Paquin et al. 2007].

CK2 phosphorylation of Puf6p is required for ASH1 mRNA localization and translation

The abrogation of Puf6p translational repression by CK2 could release the translational control of ASH1 mRNA by Puf6p. Since translation is required for proper localization of ASH1 mRNA [Chartrand et al. 2002; Irie et al. 2002], a defect in translational control by Puf6p may affect ASH1 mRNA localization. To test this, a puf6 allele with the identified CK2 phosphorylation sites (Ser31, Ser34, and Ser35) mutated to Ala was integrated into the endogenous PUF6 locus. This strain was designated SApuf6. Cell growth and morphology appeared normal. The amounts and nuclear localization of the mutant SApuf6-GFP were similar to wild-type Puf6-GFP [data not shown]. The mutant SApuf6 still interacted with Fun12p as it communoprecipitated with Tap-tagged Fun12p [data not shown]. We found that 60% ASH1 mRNA was diffusely distributed in the bud of this SApuf6 strain compared with 15% in the wild type [Fig. 6A,B]. The effect of CK2-mediated Puf6p phosphorylation on ASH1 mRNA localization was further examined in cells with deletions of CKA1 or CKA2, two genes encoding catalytic subunits of CK2. Although both catalytic subunits were synthetic lethal [Padmanabha et al. 1990], loss of either catalytic subunit alone was tolerated and cells appeared phenotypically indistinguishable from wild type [Chen-Wu et al. 1988; Padmanabha et al. 1990]. Actin structure appeared normal by phalloidin staining in those mutants [Supplemental Fig. 8]. Strikingly, both cka1 and cka2 cells showed abnormalities in ASH1 mRNA localization [Fig. 6A]. In the cka1 and cka2 strains, 56% and 72% of ASH1 mRNA were diffusely localized within the bud, respectively [Fig. 6B]. The similarity of the effect in ASH1 mRNA localization observed in SApuf6, cka1, and cka2 supports the conclusion that CK2 phosphorylation of Puf6p was required for proper localization of ASH1 mRNA at the bud cortex. As ex-
expected, the Ash1p distribution was also affected in SApuf6, cka1, and cka2 mutants with an increase in cells, with Ash1p symmetrically partitioned in daughter and mother cells compared with wild type [Fig. 6C].

To assess whether the phosphorylation of Pu6p might affect the translation of endogenous ASH1, we examined the protein levels of Ash1p in wild-type, cka1, cka2, and SApuf6 mutant strains. Ash1p levels decreased by 40% ± 10% (P < 0.05) in the SApuf6 mutant, 20% in cka1, and 60% in cka2 mutants compared with the wild-type strain [Fig. 6D]. The mRNA levels of ASH1 increased by 20% in the SApuf6 mutant strain, suggesting that the decrease in Ash1p was mainly due to translational repression in SApuf6 mutant strain. The phosphorylation status of endogenous Pu6p in cka1, cka2, and SApuf6 mutants was examined by λ ppase treatment and phosphoprotein staining. We found the endogenous Pu6p still had some phosphorylation in cka1, cka2, and SApuf6 mutants [Supplemental Fig. 9], indicating there might be other potential kinases or sites involved in phosphorylation of Pu6p.

The release of Pu6p requires coordination with the transport of ASH1 mRNA. Therefore, it is reasonable to hypothesize that CK2 phosphorylation of Pu6p occurs at the bud cortex where ASH1 mRNA is localized. To test this, we performed fluorescent in situ hybridization [FISH] in cells expressing GFP-tagged Cka1p or Cka2p. CK2 has been reported as primarily nuclear in yeast (FISH) in cells expressing GFP-tagged Cka1p or Cka2p. To test this, we performed fluorescent in situ hybridization [FISH] on cells expressing GFP-tagged Cka1p or Cka2p. CK2 has been reported as primarily nuclear in yeast (FISH) in cells expressing GFP-tagged Cka1p or Cka2p. We found that CK2 also accumulated at the bud cortex and colocalized with the ASH1 mRNA [Fig. 7A,B]. The cortical CK2 was detectable in cells with small buds that had not started expression of ASH1 mRNA [Fig. 7C], suggesting that partition of the cortical CK2 occurred prior to ASH1 mRNA localization. Our findings suggest that Pu6p could be phosphorylated at the bud cortex and released from localized ASH1 mRNA, resulting in the localized synthesis of Ash1p.

Figure 7. ASH1 mRNA colocalizes with CK2 catalytic subunits at the bud cortex. ASH1-FISH (red) on cells expressing Cka1p-GFP or Cka2p-GFP [green]. Overlay showing ASH1 mRNA, GFP from CK2 catalytic subunits, and DAPI. Representative cells were in late anaphase with ASH1 expression (A,B) and cells in earlier cell cycle stage without ASH1 expression (C). Bar, 5 μm. Arrowheads indicate bud tip.

Discussion

Pu6p suppresses ASH1 mRNA translation via Fun12p/eIF5B

Pu6p belongs to a highly conserved family of RNA-binding proteins that are involved in regulating mRNA translation and stability (Spassov and Jurecic 2003). Binding of Pu6p to the 3’UTR of ASH1 mRNA could potentially affect events occurring at the 5’ of the transcript by interacting with a general translation factor(s) [Wickens et al. 2002]. eIF5B (Fun12p in S. cerevisiae) is such a general translation factor assisting 60S ribosomal subunit joining in the translation initiation (Pestova et al. 2000). We showed that Pu6p interacts with eIF5B/Fun12p and this interaction may lead to the translational repression of ASH1 mRNA. First, the binding of Pu6p interferes with 80S ribosomal complex assembly on the ASH1 mRNA, which could result from the interference with the function of Fun12p in translation initiation. Second, Ash1p levels decrease in cells with FUN12 disruption and increase when FUN12 is overexpressed, suggesting that Fun12p is specifically required for ASH1 mRNA translation. Although identified as a general translational factor [Choi et al. 1998; Lee et al. 2002], eIF5B/Fun12p has been shown to regulate translation for many specific transcripts. Interaction of eIF5B and VASA is essential for translational activation of gurken mRNA [Carrera et al. 2000; Johnstone and Lasko 2004], one of the localized transcripts important for the embryonic development of Drosophila. Interaction of human eIF5B and HIV-1 matrix was thought to generate a pool of ribosome-free mRNA for virion packaging by repressing translation initiation [Wilson et al. 1999]. In S. cerevisiae, the poly[A]-binding protein [PABP]-mediated translational regulation of poly[A] mRNA is eIF5B/eIF5-dependent [Searfoss et al. 2001]. In addition, the Fun12p–Pu6p association was abrogated when the N-terminal region of Fun12p was truncated [data not shown]. Truncated Fun12p rescued the expression of Ash1p but not ASH1 mRNA localization [data not shown], supporting the fact that the Fun12p–Pu6p interaction was critical for proper regulation of ASH1 mRNA in vivo.

The C-terminal region and PUF domain [C536] can interact with Fun12p and bind RNA. However, this fragment of Pu6p is not sufficient to repress translation, which highlights the essential role of the N-terminal region of Pu6p. This is in contrast to Drosophila Pumilio [a PUF family member], for which expression of the RNA-binding domain is sufficient to rescue abdominal segmentation defects in pum mutant embryos [Wharton et al. 1998]. Since the interaction between Pu6p and Fun12p is dependent on RNA, the finding that the N-terminal region did not pull down Fun12p from yeast extracts does not rule out the possibility that it could interact with Fun12p in vivo. However, it is possible that the N120 region of Pu6p could potentially interfere with the function of Fun12p even if it does not directly interact with Fun12p, e.g., through interaction with other factors that would otherwise interact with Fun12p. One candidate is elF1A, which has been shown to interact
mRNAs, and phosphorylation of VASA has been linked to a down-regulation of its activity in translation of gurken mRNA (Ghabrial and Schupbach 1999). It is possible that phosphorylation of VASA would also reduce its RNA binding.

CK2 regulates cell cycle and cell polarity in S. cerevisiae (Hanna et al. 1995; Rethinaswamy et al. 1998). ASH1 mRNA localization is cell cycle-regulated and actin cytoskeleton-dependent (Bobola et al. 1996; Long et al. 1997). The phosphorylation of Puf6p by CK2 could be a temporally and spatially regulated event that may be restricted to the bud cortex where ASH1 mRNA localizes. Compatible with this hypothesis, CK2 has been identified to associate with the Arp2/3 complex (Schaerer-Brodbeck and Riezman 2003) that colocalizes with filamentous actin in highly dynamic regions of the cell cortex (Moreau et al. 1996; Winter et al. 1997). Our study showed that the two catalytic subunits of CK2 were enriched at the bud tip, supporting the hypothesis that Puf6p is phosphorylated by CK2 when it reaches the bud cortex. Interestingly, the CK2 catalytic subunits were found localized to the rough ER in mammalian cells (Faust et al. 2001) and cortical ER is also localized at the bud tip in yeast cells (Preuss et al. 1991). Our findings that the catalytic subunits of CK2 localized to the bud cortex suggest that yeast CK2 catalytic subunits might also localize to the cortical ER in yeast, which has membrane-associated ribosomes.

**Translation is required for anchoring ASH1 mRNA to the bud cortex**

Translation is important for proper localization of ASH1 mRNA. Cells overexpressing Khd1p, a translational repressor, localize ASH1 mRNA less efficiently (Irie et al. 2002). Our studies find that loss of Fun12p reduced Ash1p levels and abrogated ASH1 mRNA localization. The delocalized ASH1 mRNA had a diffusion pattern different from the she mutants (Long et al. 1997) but is a phenocopy of the atg-mutant ASH1, which is found diffusely within the bud (Irie et al. 2002). This suggests that delocalization of ASH1 mRNA in the fun12 strain may result from a deficient translation but is not a secondary effect of a transport defect. The diffusion pattern of ASH1 in fun12 cells is similar to what has been found in the cells with gene disruptions of Bud6p/Aip3p or Bni1p/She5p [Beach et al. 1999]. In those mutants, GFP-labeled ASH1 transcripts migrated to the bud but failed to be immobilized at the bud tip [Beach et al. 1999]. Therefore, our results support the model that translation is required for ASH1 mRNA anchoring. We found that SApuf6 mutant showed a similar but milder phenotype of both RNA delocalization and Ash1p levels compared with fun12 cells. This correlation between RNA delocalization and Ash1p levels confirms that ASH1 mRNA localization and translation is a well-coordinated process and suggests that the release of Puf6p for translation is important for the proper anchoring of ASH1 mRNA at the bud tips. It could also be possible that the polysomes or the nascent chains of Ash1p immobilize ASH1 mRNA at the bud cortex. Recently, it has been suggested that transla-

**Phosphorylation by CK2 releases translational control by Puf6p**

The N-terminal region of Puf6p is critical for translational repression and contains the identified CK2 phosphorylation sites. CK2 is a ubiquitous Ser/Thr protein kinase that acts as a global regulator of cellular function (Litchfield 2003; Canton and Litchfield 2006). Mutation of the identified CK2 phosphorylation sites in Puf6p significantly increases the translational repression that was corroborated by the CK2 inhibitor DMAT. Consistent with our results support the model that translation is required for anchoring ASH1 mRNA at the bud cortex that the catalytic subunits of CK2 localized to the bud tip, supporting the hypothesis that Puf6p is phosphorylated by CK2 when it reaches the bud cortex. Interestingly, the CK2 catalytic subunits were found localized to the rough ER in mammalian cells (Faust et al. 2001) and cortical ER is also localized at the bud tip in yeast cells (Preuss et al. 1991). Our findings that the catalytic subunits of CK2 localized to the bud cortex suggest that yeast CK2 catalytic subunits might also localize to the cortical ER in yeast, which has membrane-associated ribosomes.

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Expression of ASH1 occurs via specific ribosomes [Komili et al. 2007] and, if so, may help to explain the specific action of Fun12p for its translation.

In conclusion, we identified a possible mechanism of the translational repression by Puf6p and propose that the repression may be released by CK2 phosphorylation.

Our work advances the understanding of the relationship and coordination between RNA localization and translation. Premature translation due to the lack of a translational repressor can result in ASH1 mRNA delocalization, which is probably caused by interference with the transport machinery. Conversely, derepression is also required for stringent ASH1 mRNA localization, which might reflect a unique role for translation in anchoring ASH1 mRNA at the bud tip. Future work will detail the temporal and structural events of the protein–protein interactions and protein–RNA interactions that mediate this highly complex regulatory process.

Materials and methods

Yeast strains and growth media

Yeast cells were grown either in rich media or in the synthetic media lacking the nutrients indicated. Yeast strains used are listed in Table 1. Transformation was performed as described [Gietz and Woods 2002]. FUN12, PUF6, CKA1, and CKA2 genes were disrupted using an HIS disruption cassette amplified by PCR from plasmid pFA6a-His3MX6 [Wach et al. 1997]. Strains expressing Fun12-HA, Puf6-HA, Cka1-GFP, Cka2-GFP, or Fun12-Tap were obtained through insertion of an HA-HIS cassette amplified from pFA6a-3HA-His, a GFP-HIS cassette amplified from pFA6a-GFP[St57]-His [Longtine et al. 1998], or a Tap-TRP cassette amplified from pBS1479 (Rigaut et al. 1999). The strain expressing Puf6-GFP-HA was obtained by two sequential insertions of cassettes amplified from pFA6a-GFP[St57]-His and pFA6a-3HA-TRP [Longtine et al. 1998]. The strain expressing Puf6-3HA-HIS was obtained through insertion of a Puf631,34,35A-3HA-TRP cassette in the puΔ strain. This cassette was obtained by conjugating a 3xA-TRP cassette from pFA6a-3HA-TRP [Longtine et al. 1998] with the Puf631,34,35A construct derived from plasmid pET30a-Puf631,34,35A through PCR. The HA tag was fused in-frame to the C terminus of the Puf631,34,35A construct. The transformed strains were selected in appropriate synthetic medium plus dextrose (Difco). Insertion of each cassette was verified by genomic PCR.

<table>
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<th>Table 1. Yeast strains used in this study</th>
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<td>K699</td>
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cocktail tablets (Roche) and snap-frozen in liquid N2. The frozen sample was thawed at 4°C and a 30,000g supernatant was prepared. Endogenous amino acid pools and low-molecular-weight inhibitors were removed by PD-10 column (Amersham). Capped and polyadenylated transcripts for the in vitro translation assay were synthesized with Mmessage Mmachine SP6 kit (Ambion) from HindIII linearized plasmid pr-luc-E3 and pr-luc. RNA was purified by DNase I digestion, phenol/chloroform extraction, and ethanol precipitation. The integrity of the transcribed RNA was verified by electrophoresis in 1% agarose gels containing 1 µg/mL ethidium bromide. In vitro translation assays were performed as described (Iizuka et al. 1994) with the following modification. RNA reporters were heated for 10 min at 65°C and cooled for 10 min at room temperature. RNA reporters of 100 ng were preincubated with recombinant His6-Puf6 as indicated in the figure legends for 30 min at room temperature and then incubated with yeast extract in the translation buffer for 40 min at 18°C as described (Iizuka et al. 1994). For autoradiography, minus Met amino acid (Promega) was used with [35S]-Met (15 mCi/mL, Amersham). For the luciferase assay, the complete amino acid mixture (Promega) was supplied to the reaction. After incubation with extracts, the total mixture was used for the luciferase assay using a renilla substrate (Promega). All luciferase assays were performed three times and the average results are shown.

Sucrose gradient analysis
Cold and 32P-labeled transcripts were generated by SP6 polymerase-directed in vitro transcription from the Smal linearized plasmid pMiniORF-E3. 32P-labeled RNA (16 ng) was preincubated with 1 µg of full-length His6-Puf6 or the indicated components in the figure legends for 30 min at room temperature. The in vitro translation reaction was then assembled as described above and loaded onto 12 mL of 7%-47% linear sucrose gradients and subjected to centrifugation at 40,000 rpm for 2.5 h in an SW41 rotor (Beckman). Fractions (0.5 mL each) were collected from top to bottom and the A460 profile of the fractions was determined with a PharmaciaLKB Uvicord SII detector (Pharmacia) equipped with an autodensi-flow (Labconco). The radioactivity of each fraction was measured by a scintillation counter. The experiments under the condition of Pu66 incubation were repeated in 5%-25% sucrose gradient in the SW51Ti and SW41 rotors, respectively.

Co-IP, pull-down, and Western blot analysis
Cells were lysed in IP buffer (50 mM Tris at pH 7.4, 10 mM MgCl2, 100 mM NaCl, 1 mM EDTA, 10% Glycerol supplied with RNaseOUT and Protease inhibitors). Cell extracts containing 1 mg of total protein were used for co-IP and pull-down assays. For co-IPs, the yeast extract was incubated with IgG Sepharose 6 Fast Flow beads (Amersham) equilibrated in IP buffer in a volume of 200 µL for 2 h at 4°C. Beads were washed with 500 µL of buffer B (10 mM Tris at pH 8.0, 150 mM NaCl, 0.1% NP40), resuspended in SDS sample buffer, boiled, and analyzed by Western blotting. For pull-down assays, yeast extract was incubated with 946 pmol of each recombinant protein for 1 h at 4°C and pulled down by His-select HF Nickle affinity gel (Sigma). Beads were washed with 500 µL of buffer B (10 mM Tris at pH 8.0, 150 mM NaCl, 0.1% NP40), resuspended in SDS sample buffer, boiled, and analyzed by Western blotting. Western blot analyses were performed according to standard proce-
dure. IR dye-conjugated secondary antibodies [Rockland] were used at 1:1,000 dilution and detected with LI-COR Odyssey Infrared Imaging [Li-cor, Inc.]. Anti-HA [Roche, 1583816], anti-c-Myc [Molecular Probes, A-21280], anti-Pgk1 [Molecular Probes, A-6457], and anti-Tap [Open Biosystems, CAB1001] antibodies were used for detection of proteins expressed in yeast cells. Recombinant Pu6 was purified from E. coli was detected by an anti-His antibody [Invitrogen, R930-25]. Anti-Rpl3B and anti-Nop1 antibodies were kind gifts from Dr. Jonathan Warner and Dr. Tom Meier (Albert Einstein College of Medicine), respectively.

**Real-time PCR**

Yeast RNA was prepared using an RNaseasy kit according to the manufacturer's instructions [Qiagen]. Real-time PCR was performed with ASH1-, ACT1-, and PGK1-specific primers using FastStart SYBR Green Master Mix and the LightCycler instrument [Roche] as instructed by the manufacturer.

**Phosphoprotein staining, λ ppase, and RNA-binding assays**

Cell extract containing Pu6p-Tap was incubated with IgG-coated Sepharose 6 Fast Flow beads [Amersham Biosciences] equilibrated in buffer B [10 mM Tris at pH 8.0, 150 mM NaCl, 0.1% NP40] for 2 h at 4°C. The extract was purified by anti-HA antibody-coupled protein G beads. Beads were washed, resuspended in SDS sample buffer, boiled, and separated on 4%–12% SDS-PAGE gels [Invitrogen]. The gel was stained with Pro-Q Diamond Phosphoprotein Gel Stain and SYPRO Ruby [Molecular Probes] as instructed by the manufacturer. After Ruby staining, the gel was analyzed by standard Western blotting. Pepper-mintStick Phosphoprotein Molecular Weight Standard is from Molecular Probes. For the λ ppase assay, proteins affinity-purified on IgG-coupled beads were treated with 3200 U of λ ppase [New England Biolabs] for 1 h at 30°C and the eluates were analyzed by Western blotting as described above. For RNA-binding assays, the 32P-labeled RNA probe was generated by SP6 polymerase-directed in vitro transcription from the Smal linearized plasmids pMiniORF-E3 and pMiniORF-E3-M, as described above. Labeled RNA was incubated with affinity-purified proteins from yeast extract bound to IgG-coupled beads in RNA-binding solution [20 mM Tris at pH 7.4, 50 mM KCl, 3 mM MgCl2, 2 mM dithiothreitol, 5% Glycerol] for 30 min at room temperature. Protein–RNA complexes were eluted with preheated SDS sample buffer and radioactivity from the liquid phase was measured using a scintillation counter.

**In vitro phosphorylation assay**

Recombinant Pu6 was incubated with yeast extract as described in the in vitro translation assay except for the following changes. RNA reporter and the amino acid mixture were omitted from the reaction, and the ATP concentration was lowered to 0.1 mM. Where indicated, [γ-32P]-ATP or [γ-32P]-GTP (4 µCi per sample; Amersham) was used. Reactions were terminated by addition of 500 µL of binding buffer [10 mM Na-KHPO4 at pH 7.4, 1 M NaCl, 10 mM β-mercaptoethanol]. Recombinant Pu6 was added in the assay was precipitated with HIS-select HP Nickel affinity gel [Sigma] and analyzed by autoradiograph and Western blotting. Two sources of CK2 were used as indicated in the legends. Pure CK2 from sea star Pisaster ochraceus [Upstate Biotechnology] was incubated with recombinant protein for 1 h at 37°C as described [Meier 1996]. Pure CK2 from rat liver [Protein] was incubated with recombinant protein in buffer [25 mM Tris at pH 7.4, 200 mM NaCl, 10 mM MgCl2, 0.1 mM ATP] for 30 min at 37°C. Where indicated, 25 µM DMAT [EMD] was added to the yeast extract 30 min before the phosphorylation assay or supplied to the pure CK2 reaction. For mass spectrometry, 46 µg of full-length His6-Pu6 were incubated with yeast extract for 2.5 h at 18°C. His6-Pu6 was pulled down with HIS-select HP Nickel affinity gel [Sigma] and eluted with preheated SDS sample buffer. Eluted proteins and pure His6-Pu6 were separated on 4%–12% SDS-PAGE gels and stained with Coomassie blue. Gel bands containing phosphorylated and unphosphorylated His6-Pu6 were excised and subjected to MALDI-TOF at Rockefeller University.

**FISH and immunofluorescence**

Yeast cells were grown to early or mid-log phase and processed for in situ hybridization and immunofluorescence staining as described in Chartrand et al. (2000). Briefly, yeast sporeplasts were hybridized with a pool of Cy3-conjugated ASH1 RNA oligonucleotide probes [Long et al. 1997]. Cells were imaged on a BX61 upright wide-field, epifluorescence microscope. An optically sectioned three-dimensional image stack of each field was acquired with a slice spacing of 200 nm in the Z-axis with a CoolSnap HQ CCD camera [Photometrics] operated by IPLab software [BD Biosciences]. Each image stack was combined using a maximum intensity projection algorithm. Image stacks from the GFP signal were deconvolved using a classic maximum likelihood estimation algorithm in Huygens Professional [Scientific Volume Imaging] to increase the sensitivity of detection. The method of quantitative measurement on the localization of ASH1 mRNA and Ash1p has been described previously (Chartrand et al. 2002). ASH1 mRNA was classified as localized when it was predominantly in the bud tip showing a crescent localization pattern. ASH1 mRNA was classified as delocalized when it was diffusely distributed in the bud or in both mother and daughter cells.

**Statistical analysis**

The results are shown as means ± S.D. Statistical analysis was performed by the Student's t-test using SigmaPlot 9.0. Significance was accepted at P values of <0.05.

**Acknowledgments**

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